Research Article

Otodectes Cynotis Infestation in A Cat Complicated by Secondary Staphylococcus Aureus Infection: Antibiotic Sensitivity Analysis

Rajesh Kumar Verma^{1*}; Sonu Jaiswal²; Saurabh¹; Alok Singh¹; Kabeer Alam³; Utkarsh Verma⁴

¹Assiatant Professor, Department of Veterinary Clinical Complex, College of Veterinary Science and Animal Husbandry, India

²Professor & Head, Department of Veterinary Clinical Complex, College of Veterinary Science and Animal Husbandry, India

³Ph.D. Scholar, Department of Gynaecology and Obstetrics, College of Veterinary Science and Animal Husbandry, India

⁴Scholar, BVSc & AH, Internship, College of Veterinary Science and Animal Husbandry, India

*Corresponding author: Rajesh Kumar Verma

Assiatant Professor, Department of Veterinary Clinical Complex, College of Veterinary Science and Animal Husbandry, Kumarganj, Ayodhya (Uttar Pradesh) 224229, India.

Email: drrajesh16@yahoo.com

Received: September 13, 2024 Accepted: October 02, 2024 Published: October 09, 2024

Abstract

A domestic cat showed signs of ear scratching, head shaking, and waxy ear discharge for more than 45 days. The cat had been previously treated by a veterinarian, but no improvement was seen. It was then brought to the Department of Veterinary Clinical Complex at the College of Veterinary Science and Animal Husbandry, Acharya Narendra Deva University of Agriculture and Technology, Kumarganj, Ayodhya (U.P.), India. The cat was diagnosed with an ear mite infestation caused by Otodectes cynotis. A secondary bacterial infection with Staphylococcus aureus was confirmed through culture, Gram staining, and biochemical tests. Antibiotic sensitivity testing showed that the S. aureus strain was resistant to many antibiotics but sensitive to vancomycin and streptomycin, with intermediate sensitivity to gentamicin and amikacin. This report emphasizes the importance of diagnosing secondary bacterial infections in cases of ear mite infestations and the need for proper antibiotic sensitivity testing to guide effective treatment of secondary infections in otitis

Keywords: Otodectes cynotis; Staphylococcus aureus; Ear mite infestation; Antibiotic sensitivity; Otitis externa; Cat; Multidrug resistance

Introduction

Ear mites, particularly *Otodectes cynotis*, are a common cause of otitis externa in cats, leading to clinical signs such as ear scratching, head shaking, and waxy discharge [10]. While mites primarily cause irritation and inflammation in the external ear, secondary bacterial infections often complicate the clinical presentation. Opportunistic bacteria such as *Staphylococcus aureus* frequently colonize the inflamed tissue, exacerbating the infection [8]. The case described here involves the diagnosis of an *Otodectes cynotis* infestation in a cat, followed by the identification of a secondary *Staphylococcus aureus* infection. Diagnostic methods included microscopic identification of the mites, bacterial culture, Gram staining, and catalase testing. Antibiotic Sensitivity Testing (AST) was also conducted to guide effective antimicrobial therapy.

Materials and Methods

Sample Collection

A domestic short-haired cat presented with signs of otitis externa, including severe ear scratching, head shaking, and dark, waxy discharge from the ears. A sterile ear swab was collected and used for diagnostic analysis.

Otodectes cynotis Identification

The ear swab was placed in 10% potassium hydroxide (KOH) for one hour to dissolve waxy debris. The sample was then examined under a light microscope at 10x magnification, and mites were identified based on their morphological features described by Baker et al. [1], including elongated bodies and short legs.

Austin Publishing Group

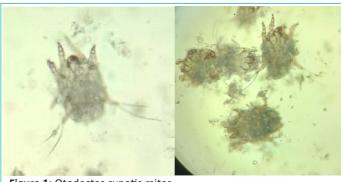


Figure 1: Otodectes cynotis mites.

Verma RK

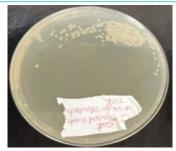


Figure 2: Growth on Nutrient Agar Plate.

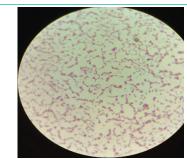


Figure 3: Gram's stain.

Bacterial Culture from Ear Swab

The same ear swab was cultured in nutrient broth for 24 hours at 37°C to promote bacterial growth. After incubation, the broth culture was streaked onto nutrient agar plates and incubated aerobically at 37°C for 24 hours. This method allows for the isolation and identification of bacterial colonies from the sample [2].

Bacterial Identification

• **Gram Staining:** Colonies grown on nutrient agar were subjected to Gram staining using standard protocols [2]. The **Table 1:** The antibiotic sensitivity results are summarized.



Figure 4: Catalase.

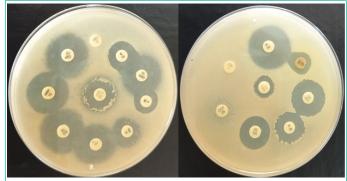


Figure 5: Antibiotic Sensitivity test.

slides were stained with crystal violet, iodine, decolorized with alcohol, and counterstained with safranin. The morphology and Gram reaction of the bacteria were observed under a microscope at 100x magnification.

• Catalase Test: A colony was placed on a glass slide, and a drop of 3% hydrogen peroxide was added. The immediate formation of bubbles (oxygen release) indicated a positive catalase reaction, suggesting the presence of *Staphylococcus aureus* [9].

Antibiotic Sensitivity Testing

The antibiotic sensitivity test was performed using the Kirby-Bauer disc diffusion method on Mueller-Hinton agar plates [4]. Eighteen antibiotic discs (Hi-Media) were placed on the Mueller-Hinton media plate, and plates were incubated at 37°C for 24 hours. Zones of inhibition were measured, and the results were interpreted according to zone size interpretation chart of Hi-media.

Results

Otodectes Cynotis Identification

Microscopic examination of the ear swab after KOH treatment confirmed the presence of *Otodectes cynotis* mites. The mites were identified based on their characteristic elongated bodies, short and stout legs, and suckers on the first and second pairs of legs.

S.No.	Antibiotics	Concentration (µg)	Diameter of Zone of Inhibition (mm)	Interpretation
	Ceftriaxone Sulbactam (CIS)	30/15	19	Resistant
	Ciprofloxacin (CIP)	10	21	Resistant
	Cotrimoxazole/Sulpha Trimethoprim (COT)	25	0	Resistant
	Vancomycin (VA)	30	18	Sensitive
	Methicillin (MET)	10	0	Resistant
	Tetracycline (TE)	30	12	Resistant
	Azithromycin (AZM)	30	20	Resistant
	Gentamicin (GEN)	10	22	Intermediate
	Amikacin (AK)	30	23	Intermediate
	Penicillin (P)	10 units	10	Resistant
	Enrofloxacin (EX)	10	21	Resistant
	Chloramphenicol (C)	50	18	Resistant
	Norfloxacin (NX)	10	18	Intermediate
	Oxacillin (OX)	1	0	Resistant
	Streptomycin (S)	10	23	Sensitive
	Amoxicillin (AMX)	10	11	Resistant
	Erythromycin (E)	15	15	Resistant
	Meropenem (MRP)	10	20	Resistant

Bacterial Culture and Identification from Ear and Nasal Swabs

• **Ear Swab Culture:** Colonies grew on nutrient agar after incubation, and Gram staining revealed Gram-positive cocci arranged in clusters, characteristic of *Staphylococcus aureus*. The catalase test was positive, confirming the bacterial identity.

Antibiotic Sensitivity Testing (ABST)

(Table 1)

Discussion

Otodectes cynotis is a prevalent cause of otitis externa in cats and can often result in secondary bacterial infections [10]. In this case, Staphylococcus aureus was identified as the secondary pathogen. Gram staining and catalase testing provided rapid confirmation of the bacterial species, which is consistent with literature on diagnostic techniques [9]. The presence of Staphylococcus aureus in both the ear and nasal swabs suggests systemic involvement or possible cross-contamination, highlighting the importance of sampling multiple sites when dealing with severe infections [3]. This finding underscores the role of thorough diagnostics in uncovering potential reservoirs of infection within the same host [6].

The antibiotic sensitivity test revealed a multidrug-resistant strain of *Staphylococcus aureus*, resistant to most antibiotics except vancomycin and streptomycin. This result aligns with recent studies demonstrating the emergence of Methicillin-Resistant *Staphylococcus Aureus* (MRSA) in veterinary medicine [5]. The sensitivity to vancomycin and streptomycin provided viable treatment options, while intermediate sensitivity to gentamicin and amikacin could support combination therapy.

The nasal swab culture showing *Staphylococcus aureus* growth further indicates that nasal colonization may contribute to the persistence of the infection, a well-documented phenomenon in humans with recurrent *S. aureus* infections [7]. Systemic antibiotic therapy was initiated based on the sensitivity profile, and the cat showed marked improvement in clinical signs following treatment.

This case underscores the importance of performing both ear and nasal swab cultures in cases of otitis with suspected secondary bacterial infections. The detection of nasal *S. aureus* colonization might have clinical implications for recurrent or difficult-to-treat infections.

Conclusion

This case highlights the diagnostic challenges and treatment complexities involved in managing *Otodectes cynotis* infestations complicated by secondary bacterial infections. The use of multiple diagnostic methods, including microscopic examination, bacterial culture, Gram staining, and antibiotic sensitivity testing, was crucial in identifying the correct pathogen and guiding effective treatment. Comprehensive testing, including nasal swabs, is recommended in severe or recurring cases to ensure that all potential sources of infection are addressed.

References

- Baker EW, Wharton GW. An Introduction to Acarology. Macmillan, New York. 1956.
- 2. Barrow GI, Feltham RKA. Cowan and Steel's Manual for the Identification of Medical Bacteria. Cambridge University Press. 2003.
- Bond R. Antimicrobial resistance in dermatology. Veterinary Dermatology. 2017; 28: 413-421.
- Clinical and Laboratory Standards Institute (CLSI). Performance Standards for Antimicrobial Susceptibility Testing. 30th ed. CLSI supplement M100. 2020.
- Couto N, Jones C. Multidrug-resistant bacteria in veterinary medicine. Journal of Veterinary Medicine. 2020; 65: 451-460.
- Hendrix CM. Laboratory Procedures for Veterinary Technicians. Elsevier Health Sciences. 2016.
- Kluytmans J, van Belkum A, Verbrugh H. Nasal carriage of Staphylococcus aureus: epidemiology, underlying mechanisms, and associated risks. Clinical Microbiology Reviews. 1997; 10: 505-520.
- 8. Paterson S. Bacterial skin disease in cats. Companion Animal. 2002; 7: 25-29.
- Quinn PJ, Markey BK, Carter ME, Donnelly WJC, Leonard FC. Veterinary Microbiology and Microbial Disease. Blackwell Science. 1994.
- Scott DW, Miller WH, Griffin CE. Small Animal Dermatology. 6th ed. W.B. Saunders. 2001.